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# Protecting coronary microvascular integrity in reperfused acute myocardial infarction to improve clinical outcome

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Acute myocardial infarction is a life-threatening condition caused by an abrupt obstruction of a coronary artery, which warrants timely reperfusion by primary percutaneous coronary intervention to salvage viable myocardium. Paradoxically, reperfusion itself often causes additional damage to cardiac cells, distinct from the ischemic damage. This form of reperfusion injury causes additional damage to cardiomyocytes, but importantly, also impairs the coronary microvascular endothelial cells. Although injury to the microvasculature contributes to poor long-term patient prognosis, relatively little work focuses on protecting the microvascular endothelial cells and endothelial barrier integrity. This review discusses how endothelial barrier function is affected in reperfused acute myocardial infarction, discusses recent studies that explored therapeutic options to preserve microvascular integrity, and suggests options for future research in this field.

**Keywords:** Acute myocardial infarction, Reperfusion injury, Microvascular injury, Endothelial barrier function, No-reflow, Cardioprotection

### Introduction

Acute myocardial infarction (AMI) arises from an immediate epicardial coronary artery occlusion, which causes irreversible injury starting in the subendocardium and expanding toward the epicardium with ongoing occlusion (Reimer et al., 1977). Therefore, AMI requires timely restoration of blood flow by percutaneous coronary intervention (PCI) to salvage the myocardium (Ibanez et al., 2018). Paradoxically, in approximately half of PCI-treated AMI patients (van Kranenburg et al., 2014; de Waha et al., 2017), reperfusion of previously ischemic tissue inflicts incremental cardiac damage distinct from ischemic damage, hampering restoration of myocardial perfusion. This form of ischemia-reperfusion (IR) injury affects the coronary microvasculature and is known as microvascular obstruction or microvascular injury (MVI) (Robbers et al., 2013; Hollander et al., 2016; Konijnenberg et al., 2020). Independent of infarct size, MVI contributes to worse patient prognosis (van Kranenburg et al., 2014; de Waha et al., 2017). MVI is characterized by a cascade of pathophysiological changes, including loss of microvascular integrity, endothelial cell dysfunction, endothelial cell damage, and the development

of intramyocardial hemorrhage (IMH) (Robbers et al., 2013; Hollander et al., 2016; Nair et al., 2020; Liu et al., 2022). This highlights that targeting MVI, in addition to current care, may represent a clinically relevant area of study. Although currently no widely approved therapeutic options for MVI exists in AMI patients, some preclinical studies have reported promising results. Targeting coronary endothelial barrier function, with the purpose of limiting the extent of MVI, could be a promising adjuvant strategy in addition to current care to improve patient outcome. This review discusses how microvascular endothelial cells are affected in reperfused AMI, discusses recent (pre) clinical studies on therapeutic options on top of current care to attenuate MVI, and highlights future directions.

### The coronary microvasculature

The coronary microvasculature is a heterogenous network of vessels with a diameter smaller than 100  $\mu\text{m}$ , including arterioles, capillaries, and venules. The microvasculature adapts to physiological and pathophysiological stimuli and has a key role in regulating vascular tone, blood flow, and oxygen transport. Blood that enters the coronary microcirculation starts

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in arterioles, which consist of endothelial cells and basement membrane, and are surrounded by vascular smooth muscle cells. Arterioles are the main regulators of myocardial blood flow. Subsequently, blood flows into the capillaries, which consist of a single layer of endothelial cells surrounded by basement membrane and are closest to the cardiomyocytes. Capillaries are crucial in the exchange of nutrients, oxygen, and waste products to the cardiomyocytes. Finally, capillaries converge in postcapillary venules that allow deoxygenated blood return to the venous system (Fonseca et al., 2016).

In the context of reperfusion-induced microvascular injury, several components of the coronary capillaries are important to highlight: the glycocalyx, the endothelial cell and its cell junctions, and the basement membrane. The glycocalyx covers the apical surface of the endothelial cell and consists of a network of mainly glycoproteins, proteoglycans, and plasma and endothelium-derived soluble components. The glycocalyx acts as the first line of defense for the cell against MVI (Reitsma et al., 2007). Endothelial cells line the interior surface of the vessel and are connected by cell-cell junctions, including adherens junctions and tight junctions, which permit passage of water and other molecules across the endothelium (Komarova et al., 2017). These cell-cell junctions form a crucial part in maintaining and preserving the integrity of the microvasculature. The basement membrane is attached to the basolateral side of the endothelium and consists of networks of mainly laminin and type IV collagen (Jayadev and Sherwood, 2017).

#### Determinants of microvascular injury during reperfusion

During myocardial ischemia, oxygen transport to the myocardium and endothelial cells is significantly restricted. Prolonged ischemia results in reduced cytosolic adenosine triphosphate production, affects ion-exchange channels, and induces anaerobic metabolism and metabolic acidosis (Pike et al., 1993). In experimental models, a longer duration of ischemia seemed to be an important determinant of the severity of MVI to a large extent (Kloner et al., 1974; Reffellmann et al., 2002). Clinical trials, however, show conflicting results regarding the association between time to reperfusion and MVI severity (Husser et al., 2013; Amier et al., 2017; Ferré-Vallverdú et al., 2021). One potential reason is that endothelial cells are relatively resistant to hypoxia (Quintero et al., 2006; Baldea et al., 2018). Anaerobic glycolysis can provide sufficient energy for endothelial cells (Culic et al., 1997), making them less prone to hypoxic injury compared to cardiomyocytes (Mertens et al., 1990). Although endothelial cells tolerate hypoxic conditions relatively well, endothelial cells are particularly vulnerable to reperfusion injury (Maxwell and Gavin, 1991; Hollander et al., 2016). Indeed, reperfusion results in an immediate, progressive deterioration of microvascular integrity (Hollander et al., 2016; Sezer et al., 2022), underlining the importance of early intervention. Upon presence of IR injury, key findings at the ultrastructural level include occurrence of swelling and blebbing of endothelial cells, presence of membrane-bound vesicles and cellular debris in the perivascular space (Maxwell and Gavin, 1991). In more severe cases, reperfusion injury causes thinning and rupture of endothelial cells, reduction in endothelial cell-junctions, and extravasation of erythrocytes (Hollander et al., 2016), whereas ischemia alone induces only mild morphological changes to the coronary microcirculation (Maxwell and Gavin, 1991; Hollander et al., 2016). These changes in endothelial cell function and structure upon reperfusion lead to increased vascular permeability and additional cellular damage. Mechanistically, various components modulate the extent of vascular permeability (Figure 1). Identifying the most important factors within this process could result in the development of novel therapeutic approaches, specifically targeted at these

factors, in an attempt to further limit MVI (Table 1).

#### Strategies to minimize coronary microvascular injury

Prevention of MVI may have strong clinical potential, especially since MVI is related to clinical outcomes and MVI is currently not directly targeted. For this purpose, we have summarized studies that focused on potential targets of MVI; i.e. gradual reperfusion, oxidative stress, glycocalyx, endothelial barrier, and basement membrane.

#### *Mechanical reperfusion by primary percutaneous coronary intervention*

Primary PCI causes an abrupt restoration of intracoronary blood flow and pressure, leading to an imbalance between hydrostatic and colloid osmotic forces, and consequently increased vascular permeability and intramyocardial edema. Gradual or staged reperfusion could be an alternative approach to preserve coronary microvascular integrity. Gradual reperfusion has shown favorable effects in various experimental models (Okamoto et al., 1986; Takeo et al., 1995; Sato et al., 1997; Bopassa et al., 2005; Musiolik et al., 2010; Ferrera et al., 2015), including reduction in final myocardial infarct size (Okamoto et al., 1986; Sato et al., 1997; Bopassa et al., 2005; Musiolik et al., 2010; Ferrera et al., 2015) and better-preserved microvascular integrity (Okamoto et al., 1986). Several cardioprotective mechanisms of gradual reperfusion have been proposed; reduction of reactive oxygen species (ROS) production (Bopassa et al., 2005), preservation of endothelial function (Sato et al., 1997), and lowering reperfusion-induced sodium and calcium overload (Takeo et al., 1995). In contrast to the promising effects in preclinical work, gradual reperfusion in clinical trials has shown mixed results (Carrick et al., 2014; Kelbæk et al., 2016; Sezer et al., 2022). In patients with ST-elevation myocardial infarction (STEMI), gradual reperfusion by deferred stent implantation approximately nine hours after the index procedure reduced the extent of no-reflow and increased myocardial salvage at six month follow-up (Carrick et al., 2014). In contrast, deferred stent implantation approximately three days after the index procedure did not show any improvement in patient prognosis at the approximately 3.5 year follow-up (Kelbæk et al., 2016). Interestingly, a recent pilot study showed that pressure-controlled, gradual reopening of the culprit artery with deferred stent implantation 30 minutes after initial reperfusion resulted in improved microcirculatory response to reperfusion and enzymatic smaller myocardial infarct size compared to PCI with intermediate stenting (Sezer et al., 2022). Being in its infancy, clinical studies that aim to reduce MVI through (pressure-controlled) gradual reopening of the infarct-related artery could have benefit over immediate stenting. We suggest future work is required, including larger sized studies with longer follow-up, but also work around improving strategies around reopening the infarct-related artery.

#### *Oxidative stress*

ROS are generated at high levels by the re-introduction of oxygen. The role of ROS in reperfusion-induced microvascular dysfunction has been extensively reviewed by Yu et al (2019). Sources of ROS production include uncoupled endothelial nitric oxide synthase (eNOS), mitochondria, infiltrating leukocytes, and vascular adhesion protein-1. During reperfusion, ROS levels can exceed the antioxidant defense system, resulting in cellular oxidative stress that aggravates MVI. Moreover, increased ROS generation can directly and indirectly lower intracellular nitric oxide (NO) bioavailability. Increased production of the reactive oxygen ion superoxide during reperfusion interacts with NO to form peroxynitrate, in turn inhibiting endothelial-dependent vasorelaxation (O'Donnell et al., 1997). Furthermore, ROS can indirectly lower NO levels by uncoupling eNOS, which modifies eNOS monomers into ROS-

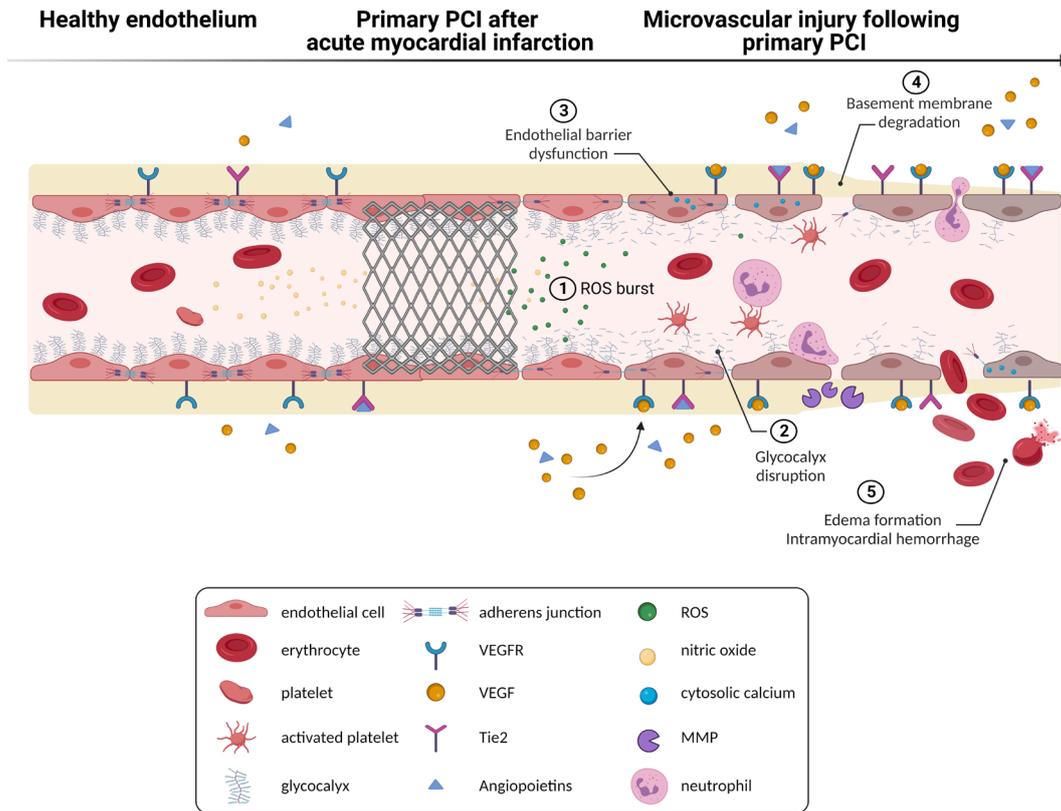


Figure 1. Reperfusion of an occluded coronary artery is often accompanied by microvascular injury. Microvascular injury is reflected by a sequelae of pathophysiological changes, including increased production of reactive oxygen species, disruption of the glycocalyx, endothelial barrier dysfunction with increased cytosolic calcium levels, loss of endothelial cell-cell junctions, degradation of the basement membrane, and in more severe cases formation of edema and intramyocardial hemorrhage. In turn, hemolysis of erythrocytes can expose cardiomyocytes to cytotoxic heme. Preclinical studies suggest that pharmacologically targeting components of the coronary microvasculature may have potential as an adjuvant therapy in acute myocardial infarction patients. MMP, matrix metalloproteinases; PCI, percutaneous coronary intervention; ROS, reactive oxygen species; Tie2, receptor tyrosine kinase; VEGF, vascular endothelial growth factor; VEGFR, VEGF-receptor.

producing units (Landmesser et al., 2003). However, clinical trials that administered vasodilating agents did not show a reduction in MVI (Vijayalakshmi et al., 2006; Desmet et al., 2011; Nazir et al., 2016).

Studies that target ROS formation, including antioxidant treatment and various forms of preconditioning, have mainly focussed on cardiomyocyte protection and have led to contradictory results (Rodrigo et al., 2013; Kalogeris et al., 2014; Zhou et al., 2015; Rodrigo et al., 2022). In part, these conflicting results may relate to the observation that, albeit ROS is involved in damage to the microvasculature, at certain dosages ROS has cardioprotective effects. Studies that specifically aim to reduce reperfusion-induced endothelial cell damage are scarce. In a mouse model of cerebral IR injury, transgenic overexpression of antioxidant superoxide dismutase (SOD) showed less vascular endothelial cell death, but did not reduce the rate of hemorrhagic transformation (Maier et al., 2006). In isolated rat hearts, SOD did not preserve coronary vascular reactivity nor improve left ventricular function after IR (Whitman et al., 1997). However, in isolated rat hearts subjected to IR, treatment with Coenzyme Q10 (CoQ10), which is a component of the mitochondrial electron transport chain, preserved endothelium-dependent and -independent vasodilation via free radical scavenger action, and improved left ventricular function (Whitman et al., 1997). Additionally, CoQ10 treatment reduced infarct size in rats, guinea pigs, and Yorkshire pigs (Awad et al., 2022). This means that targeting ROS shows promise in limiting endothelial cell injury in preclinical studies, but no clinical data is available yet. In addition, the ability of ROS (dependent on its dose) to be both harmful and protective

against injury, makes ROS a target in this area challenging.

### Glycocalyx disruption

A 0.5  $\mu\text{m}$  thick glycocalyx layer covers the vascular endothelium (van den Berg et al., 2003; Nieuwdorp et al., 2008) and consists of mainly proteoglycans (syndecans and glypicans) with long glycosaminoglycan side chains (such as heparan sulfate and hyaluronan), glycoproteins with short carbohydrate side chains, and soluble components such as albumin, thrombomodulin, and extracellular SOD (Reitsma et al., 2007). The endothelial glycocalyx plays a crucial role in vascular permeability (Vink and Duling, 2000; van Haaren et al., 2003; Rehm et al., 2004; Betteridge et al., 2017) and shear stress-induced signalling pathways, including NO production (Pahakis et al., 2007; Green et al., 2017). In animal experiments, enzymatic degradation of the glycocalyx resulted in an increase of the interstitial space, indicative of myocardial edema (van den Berg et al., 2003). In isolated guinea pig hearts subjected to short IR, addition of enzymatic degradation of the glycocalyx significantly increased vascular permeability (Rehm et al., 2004). Ischemia induces only limited damage to the glycocalyx, whereas addition of reperfusion strongly accelerates the damage to the glycocalyx (Beresewicz et al., 1998). Shedding of endothelial glycocalyx has been reported in experimental IR models (Beresewicz et al., 1998; Platts et al., 2003) and in patients undergoing cardiac surgery such as aortic valve replacement (Bruegger et al., 2015; Passov et al., 2021). The consistency of the glycocalyx can be modulated by several factors, including shear stress (Wang et al., 2020), locally produced ROS (Rubio-Gayosso et al., 2006) and cytokines

**Table 1. Approaches that primarily target components of the coronary microvasculature**

| Target  | Treatment   | Model   | Key finding(s)  |
|---|---|---|---|
| <b>1. Mechanical reperfusion</b>                      | Pressure-controlled, gradual reperfusion (Sezer et al., 2022)                                 | ST-elevation myocardial infarction patients, IR   | ↑ coronary microvascular integrity<br>↓ enzymatic infarct size  |
| <b>2. Oxidative stress</b>                            | Superoxide Dismutase (Whitman et al., 1997)   | Isolated rat heart, IR  | = coronary vascular reactivity<br>= left ventricular function   |
|   | Coenzyme Q10 (Whitman et al., 1997; Awad et al., 2022)  | Isolated rat heart, IR<br><br>Rats, guinea pigs, Yorkshire pigs, IR                           | ↑ endothelium-dependent and -independent vasodilation<br>↑ left ventricular function<br><br>↓ infarct size<br>↑ myocardial function |
| <b>3. Glycocalyx</b>                                  | Exogenous Nitric Oxide (Bruegger et al., 2008)  | Isolated guinea-pig heart, IR   | ↓ coronary vascular resistance<br>↓ fluid extravasation<br>↓ tissue oedema  |
|   | Superoxide Dismutase (Beresewicz et al., 1998)  | Isolated guinea-pig heart, IR   | ↑ endothelium-dependent vasodilation  |
|   | Hydrocortisone (Chappell et al., 2007; Chappell et al., 2010)                                 | Isolated guinea-pig heart, IR   | ↓ transudate formation  |
|   |   | Isolated guinea-pig heart, IR   | ↓ neutrophil adhesion   |
|   | Sevoflurane (Annecke et al., 2010; Chappell et al., 2011; Chen et al., 2016)                  | Isolated guinea-pig heart, IR   | ↓ transudate formation  |
|   |   | Isolated guinea-pig heart, IR<br><br>Isolated guinea-pig heart, IR                            | ↓ cell adhesion<br><br>↓ coronary vascular permeability   |
| Specific deletion of Angiotensin-2 (Lee et al., 2018) | Mouse, IR   | ↓ vascular leakage<br>↓ infarct size  |   |
| <b>4. Cell-junctions</b>                              | Relaxin (Gao et al., 2019)  | Mouse, IR   | ↑ VE-cadherin expression<br>↓ no-reflow<br>↓ vascular leakage<br>= infarct size<br>↑ myocardial function                            |
|   | Losartan (Li et al., 2019)  | Mouse, IR   | ↑ VEGFR2-Src-VE-cadherin complex formation<br>↓ vascular permeability<br>↓ haemorrhage<br>↓ infarct size                            |
|   | Angiotensin-like 4 knock-out (Galaup et al., 2012)  | Mouse, IR   | ↓ VEGFR2/VE-cadherin complex formation<br>↑ vascular permeability<br>↑ haemorrhage<br>↑ no-reflow<br>↑ infarct size                 |
|   | Human-recombinant Angiotensin like-4 (Galaup et al., 2012)                                    | Rabbit, IR  | ↓ no-reflow<br>↓ haemorrhage<br>↓ infarct size  |
|   | Endothelial-specific overexpression of Histidine triad nucleotide-binding 2 (Li et al., 2021) | Mouse, IR   | ↑ VE-Cadherin/β-Catenin interaction<br>↓ microvascular leakage<br>↓ no-reflow<br>↓ infarct size                                     |
|   | Angiotensin-1 (Lee et al., 2011)  | Mouse, IR   | ↓ VE-cadherin phosphorylation<br>↓ microvascular leakage<br>↓ infarct size<br>↑ myocardial function                                 |
|   | Imatinib (Hamid et al., 2022)   | Mouse, non-reperused myocardial infarction  | ↓ cardiac remodelling, including:<br>↑ capillary density<br>↓ fibrosis<br>↓ inflammation  |
|   | <b>5. Basement membrane</b>   | Matrix metalloproteinase-inhibitory nitrate hybrid compound (Krzywonos-Zawadzka et al., 2019) | Isolated rat heart, IR  |

IR, ischemia-reperfusion model; VE-cadherin, vascular endothelial cadherin; VEGFR2, vascular endothelial growth factor receptor 2

(Wiesinger et al., 2013). Thickness and stiffness of the glycocalyx is reduced by tumor necrosis factor- $\alpha$  (Wiesinger et al., 2013) via activation of matrix metalloproteinases (MMPs) (Ramnath et al., 2014). Furthermore, degradation and shedding of the glycocalyx contribute to leukocyte-endothelial cell adhesion (Constantinescu et al., 2003; Schmidt et al., 2012).

Several studies in animals have focussed primarily on the preservation of the glycocalyx after IR and its potential clinical benefit. In isolated guinea pig hearts, NO reduced shedding of the glycocalyx, accompanied by reduced coronary vascular resistance, coronary fluid extravasation, and tissue edema

(Bruegger et al., 2008). Also SOD (Beresewicz et al., 1998; Rubio-Gayosso et al., 2006), hydrocortisone (Chappell et al., 2007; Chappell et al., 2010), and sevoflurane (Annecke et al., 2010; Chappell et al., 2011; Chen et al., 2016) have shown to preserve the endothelial glycocalyx after IR. In mice subjected to IR, endothelial specific deletion of angiotensin-2 attenuated endothelial glycocalyx degradation and vascular leakage, and reduced infarct size in the acute phase of myocardial infarction (Lee et al., 2018). Interestingly, simulating a new glycocalyx by a selectin-targeting glycocalyx-mimetic reduced neutrophil extravasation, macrophage accumulation, improved cardiac

function, and reduced infarct size (Dehghani et al., 2022). These preclinical studies suggest that preventing glycocalyx degradation in IR can be a promising goal in cardioprotection, although studies in humans are lacking to date.

### ***Endothelial barrier dysfunction***

Endothelial cells form the inner lining of the vascular system and maintain vascular homeostasis. An early indicator of loss of vascular integrity is an increase in vascular permeability, which is mainly controlled by endothelial cell calcium homeostasis and cell junctions, importantly contributing to endothelial dysfunction.

### ***Calcium homeostasis***

Calcium ( $\text{Ca}^{2+}$ ) is a second messenger involved in signalling pathways that are crucial for regulating endothelial barrier function. Reperfusion can provoke endothelial barrier dysfunction by causing an increase in cytosolic  $\text{Ca}^{2+}$  levels and changing endothelial cell shape by activating its cytoskeletal contractile elements. Consequently, the formation of intercellular gaps promotes endothelial permeability and edema formation (Kasseckert et al., 2009; Li et al., 2020). Furthermore,  $\text{Ca}^{2+}$  plays a critical role in cell-cell adhesion. Chelation of extracellular  $\text{Ca}^{2+}$  promotes the internalization of the cell adhesive cadherin molecules (Le et al., 1999). On the other hand, repletion of extracellular  $\text{Ca}^{2+}$  resulted in restoration of vascular endothelial (VE)-cadherin integrity (Gao et al., 2000).

### ***Endothelial cell junction disruption***

Endothelial permeability is regulated via transcellular (through the endothelial cell via caveolae-mediated vesicular transport) and paracellular (between the endothelial cells via interendothelial cell junctions) pathways. Here we will focus on the paracellular pathway since this is the dominant mechanism of increased vascular permeability in the heart under pathophysiological conditions (Bazzoni and Dejana, 2004). In endothelial cells, adherens junctions and tight junctions are intermingled along sites of cell-to-cell contacts and have crucial roles in cell-signaling (Bazzoni and Dejana, 2004; Dejana, 2004; Hu et al., 2013).

### ***Adherens junctions***

The key component of adherens junctions is the transmembrane adhesion protein VE-cadherin. VE-cadherin interacts with intracellular proteins, including  $\beta$ - and  $\gamma$ -catenins. In turn, these catenins link to  $\alpha$ -catenin and connect VE-cadherin to actin filaments, supporting junctional stability (Bazzoni and Dejana, 2004; Dejana, 2004; Hu et al., 2013). The function of VE-cadherin and catenins can be modulated by tyrosine phosphorylation (Esser et al., 1998; Potter et al., 2005; Orsenigo et al., 2012; Wessel et al., 2014). For example, VE-cadherin phosphorylation at the Tyr<sup>658</sup> residue was shown to disrupt endothelial barrier function in vitro (Potter et al., 2005). In vivo, besides the VE-cadherin Tyr<sup>658</sup> residue, the Tyr<sup>685</sup> residue seems to play an important role in endothelial barrier function. In response to mediators such as vascular endothelial growth factor (VEGF), histamine, and bradykinin, phosphorylated VE-cadherin is internalized and ubiquitinated, worsening endothelial barrier function (Orsenigo et al., 2012; Wessel et al., 2014).

### ***Tight junctions***

Key components of tight junctions are claudin, occludin, and junction adhesion molecules (JAMs) (Bazzoni and Dejana, 2004; Dejana, 2004; Hu et al., 2013). Of the claudin members, endothelial cells predominantly express claudin-5. In claudin-5 deficient mice, blood-brain barrier permeability was increased for small molecules, suggesting that claudin-5 actively contributes to maintaining endothelial barrier function (Nitta et al., 2003). Claudin, occludin and JAMs link with the actin cytoskeleton via zona occludin-1 protein, thereby stabilizing

endothelial barrier function (Bazzoni and Dejana, 2004; Dejana, 2004; Hu et al., 2013). Besides its role in regulating endothelial barrier function, JAMs appear to mediate direct leukocyte/platelet/endothelial cell binding interactions (Garrido-Urbani et al., 2014).

### ***Targeting endothelial cell junctions***

Function of both adherens and tight junctions are important in maintaining vascular integrity. Whereas ischemia alone did not reduce the number of endothelial cell junctions in a rat coronary artery model, subsequent reperfusion disrupted almost all cell junctions (Hollander et al., 2016). This further highlights the importance of the reperfusion phase in causing injury to the (coronary) microvasculature. In a mouse model of myocardial IR injury, the peptide hormone relaxin partially restored VE-cadherin protein expression, and subsequently reduced the extent of no-reflow, and improved myocardial function (Gao et al., 2019). Furthermore, losartan, an angiotensin II receptor blocker, inhibited phosphorylation of Src and VE-cadherin. This resulted in increased VEGFR2-Src-VE-cadherin complex formation, a reduction in vascular permeability, hemorrhage, and infarct size (Li et al., 2019). Moreover, in angiotensin-like 4 (ANGPLT4) knock-out mice, vascular permeability, edema, hemorrhage, no-reflow, and infarct size were increased, accompanied by dissociation of the VEGFR2/VE-cadherin complex (Galaup et al., 2012). Injecting recombinant ANGPTL4 in rabbits reduced the extent of hemorrhage, no-reflow, and infarct size (Galaup et al., 2012). In another study, overexpression of histidine triad nucleotide-binding 2 enhanced VE-cadherin/ $\beta$ -catenin interaction, and subsequently attenuated microvascular leakage, improved myocardial perfusion, and reduced infarct size (Li et al., 2021). Angiotensin-1, an endothelial-specific angiogenic factor and ligand for the endothelial-specific Tie2 receptor, promoted endothelial barrier function via reduction of tyrosine phosphorylation of VE-cadherin, reduced infarct size, and improved myocardial function (Lee et al., 2011). In addition to these targeted preclinical studies, several studies explored the impact of the tyrosine kinase inhibitor imatinib, and found it to be effective in preserving endothelial barrier function in cultured endothelial cells, mainly by interfering with VE-cadherin and  $\beta$ -catenin (Dejana et al., 2008; Aman et al., 2012; Chislock and Pendergast, 2013). Interestingly, imatinib has been investigated in a wide range of animal models. In a sepsis model (Aman et al., 2012), pulmonary IR model (Tanaka et al., 2016; Magruder et al., 2018), cardiopulmonary bypass model (Koning et al., 2018), and cerebral IR models (Su et al., 2008; Merali et al., 2015), imatinib was able to reduce the extent of microvascular leakage. In addition, in mice with nonreperfused myocardial infarction, treatment with imatinib alleviated cardiac remodeling, including a higher capillary density, less fibrosis, and less inflammation compared to vehicle (Hamid et al., 2022).

Taken together, these preclinical studies provide support for pharmacologically targeting endothelial cell-cell junctions in order to prevent injury to the endothelial barrier and limit reperfusion-induced microvascular leakage. These effects may ultimately translate to smaller infarct sizes, such as demonstrated in preclinical studies, although studies in humans are currently lacking.

### ***Basement membrane degradation***

The endothelial basement membrane is a thin, continuous layer of extracellular matrix and has a critical role in vessel stabilization. A basement membrane mainly consists of collagens, laminins, and proteoglycans, and can be degraded by proteinases, such as MMPs (Davis and Senger, 2005). Neutrophils are a major source of MMPs (Romanic et al., 2002). Of those MMPs, MMP-2 and MMP-9 play an important role in different forms of IR injury (Cheung et al., 2000;

Peterson et al., 2000; Romanic et al., 2002; Sumii and Lo, 2002; Machado et al., 2006). Therefore, MMPs represent a potential target in IR, and inhibitors of MMPs hold promise as protection against MVI. Indeed, in a cerebral IR model, rats treated with a broad spectrum MMP inhibitor showed significantly reduced hemorrhagic volumes and mortality rate (Sumii and Lo, 2002). In support of these observations specific inhibition of MMP-9 also reduced cerebral hemorrhage, swelling, infarction, and mortality (Saleem et al., 2021). Comparable observations were reported in isolated rat hearts, as treatment with a MMP-inhibitory nitrate hybrid compound increased coronary flow and improved myocardial function after IR (Krzywonos-Zawadzka et al., 2019). In summary, these preclinical studies provide support for using (selective) MMP-inhibitors to attenuate injury to the basement membrane following reperfusion and may translate to (partial) preservation of myocardial function following IR.

### Current challenges in translation from bench to bedside

Despite numerous experimental animal studies with promising cardioprotective strategies, the translation into clinical therapy is challenging and has been disappointing. As already extensively reviewed (Heusch, 2017; Kleinbongard et al., 2020), several experimental specific factors, such as study design and reproducibility, and animal specific factors, such as the lack of co-morbidities and the use of co-medication, have significantly contributed to the loss of translation in cardioprotection from bench to bedside. Indeed, in most preclinical studies young and healthy animals have been used, but mainly with the purpose to reveal underlying mechanisms rather than demonstrating translational value. Regarding AMI models, animal gender should be thoroughly reported, as estrogen can be a major driver of microvascular cardioprotective pathways (Querio et al., 2021). These specific translational issues have already been addressed by Bøtker and colleagues, who published practical guidelines to ensure rigor and reproducibility of experiments in cardioprotection, including important remarks on experimental design and predefined in- and exclusion criteria (Bøtker et al., 2018). Furthermore, Lecour and colleagues (2021) published step-by-step criteria to improve the likelihood of translation of cardioprotective interventions into clinical therapy. Both guidelines will facilitate in designing more robust future cardioprotective approaches.

To date, in the clinical setting, no risk factors or clinical predictors have consistently been found for the development of MVI and/or IMH following AMI. Besides the use of additional glycoprotein IIb/IIIa inhibitors (Amier et al., 2017) and the anterior infarct location (Amier et al., 2017; Reinstadler et al., 2019), it remains unclear why some patients show MVI while others do not. Some evidence exists that admission glucose level in STEMI patients is independently related to MVI (Jensen et al., 2011) and IMH (Ota et al., 2022). However, having diabetes (Amier et al., 2017; Ota et al., 2022) or lowering blood glucose concentration with exenatide, a glucagon-like peptide-1 analogue, did not affect MVI (Roos et al., 2016). Interestingly, one day after primary PCI for STEMI, serum syndecan-1 level, a major component of the endothelial glycocalyx, was independently associated with MVO (Huang et al., 2021). Furthermore, local MMP-9 levels in the coronary artery upon reperfusion was higher in patients with IMH (Ota et al., 2022), showing the potential of coronary microvascular degradation products as biomarker for reperfusion injury over clinical risk factors.

In addition to optimizing the preclinical study design according to the consensus guidelines (Bøtker et al., 2018; Lindsey et al., 2018; Lecour et al., 2021), patient selection should be improved. Focusing on MVI, patients with a relatively short ischemic time, a large area at risk (such as can

be expected in anterior infarcts), and a completely occluded coronary artery prior to primary PCI will benefit the most from cardioprotective strategies (Heusch, 2017). Importantly, as the first few minutes to hours is the most critical phase in generating reperfusion injury, timing of pharmacological interventions should be prior to or immediately at the onset of reperfusion, in addition to current care.

To the best of our knowledge, no clinical trials primarily targeting components of the coronary microvasculature exist. Changing focus from primarily targeting cardiomyocytes to the coronary microvasculature could pave the way for novel therapeutic approaches in reperfusion injury, starting in rodent models and proceeding to the porcine model, as this most closely resembles the (patho)physiology of the human heart (Heusch et al., 2011).

### Conclusion

Microvascular barrier function requires intact glycocalyx, endothelium, and basement membrane. Damage to any of these vascular components, which seems a common feature in IR injury, can result in a leaky microcirculatory environment, edema formation, hemorrhage, and impaired ventricular function. Accordingly, microvascular injury in AMI patients seems a potential target, especially since current medicine does not directly target microvascular injury, but also because microvascular injury predicts clinical outcomes independent of infarct size. While studies in AMI patients on minimizing reperfusion injury have demonstrated limited clinical benefit, preclinical studies suggest that pharmacologically targeting the glycocalyx, the endothelium, endothelial cell junctions and/or the basement membrane may have potential as an adjuvant therapy in AMI patients. Based on these preclinical studies, primarily targeting endothelial cell-junctions seems the most promising approach in limiting microvascular injury and subsequent myocardial damage. Future work is required, especially in translating these observations to humans, to better understand the potential benefits of targeting the microvasculature to further minimize injury following AMI.

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### References

- Aman J, van Bezu J, Damanafshan A, Huveneers S, Eringa EC, Vogel SM, Groeneveld AB, Vonk Noordegraaf A, van Hinsbergh VW, van Nieuw Amerongen GP (2012) Effective treatment of edema and endothelial barrier dysfunction with imatinib. *Circulation* 126:2728-2738.
- Amier RP, Tijssen RYG, Teunissen PFA, Fernández-Jiménez R, Pizarro G, García-Lunar I, Bastante T, van de Ven PM, Beek AM, Smulders MW, Bekkers S, van Royen N, Ibanez B, Nijveldt R (2017) Predictors of Intramyocardial Hemorrhage After Reperfused ST-Segment Elevation Myocardial Infarction. *J Am Heart Assoc* 6:e005651.
- Annecke T, Chappell D, Chen C, Jacob M, Welsch U, Sommerhoff CP, Rehm M, Conzen PF, Becker BF (2010) Sevoflurane preserves the endothelial glycocalyx against ischaemia-reperfusion injury. *Br J Anaesth* 104:414-421.
- Awad K, Sayed A, Banach M (2022) Coenzyme Q(10) Reduces Infarct Size in Animal Models of Myocardial Ischemia-Reperfusion Injury: A Meta-Analysis and Summary of Underlying Mechanisms. *Front Cardiovasc Med* 9.

- Baldea I, Teacoe I, Olteanu DE, Vaida-Voievod C, Clichici A, Sirbu A, Filip GA, Clichici S (2018) Effects of different hypoxia degrees on endothelial cell cultures-Time course study. *Mech Ageing Dev* 172:45-50.
- Bazzoni G, Dejana E (2004) Endothelial cell-to-cell junctions: molecular organization and role in vascular homeostasis. *Physiological reviews* 84:869-901.
- Beresewicz A, Czarnowska E, Maczewski M (1998) Ischemic preconditioning and superoxide dismutase protect against endothelial dysfunction and endothelium glycocalyx disruption in the postischemic guinea-pig hearts. *Mol Cell Biochem* 186:87-97.
- Betteridge KB, Arkill KP, Neal CR, Harper SJ, Foster RR, Satchell SC, Bates DO, Salmon AHJ (2017) Sialic acids regulate microvessel permeability, revealed by novel in vivo studies of endothelial glycocalyx structure and function. *The Journal of physiology* 595:5015-5035.
- Bopassa JC, Michel P, Gateau-Roesch O, Ovize M, Ferrera R (2005) Low-pressure reperfusion alters mitochondrial permeability transition. *Am J Physiol Heart Circ Physiol* 288:2750-2755.
- Bøtker HE, Hausenloy D, Andreadou I, Antonucci S, Boengler K, Davidson SM, Deshwal S, Devaux Y, Di Lisa F, Di Sante M, Efentakis P, Femminò S, García-Dorado D, Giricz Z, Ibanez B, Iliodromitis E, Kaludercic N, Kleinbongard P, Neuhäuser M, Ovize M, Pagliaro P, Rahbek-Schmidt M, Ruiz-Meana M, Schlüter KD, Schulz R, Skyschally A, Wilder C, Yellon DM, Ferdinandy P, Heusch G (2018) Practical guidelines for rigor and reproducibility in preclinical and clinical studies on cardioprotection. *Basic Res Cardiol* 113:39.
- Bruegger D, Rehm M, Jacob M, Chappell D, Stoeckelhuber M, Welsch U, Conzen P, Becker BF (2008) Exogenous nitric oxide requires an endothelial glycocalyx to prevent postischemic coronary vascular leak in guinea pig hearts. *Critical care (London, England)* 12:R73.
- Bruegger D, Brettner F, Rossberg I, Nussbaum C, Kowalski C, Januszewska K, Becker BF, Chappell D (2015) Acute degradation of the endothelial glycocalyx in infants undergoing cardiac surgical procedures. *Ann Thorac Surg* 99:926-931.
- Carrick D, Oldroyd KG, McEntegart M, Haig C, Petrie MC, Eteiba H, Hood S, Owens C, Watkins S, Layland J, Lindsay M, Peat E, Rae A, Behan M, Sood A, Hillis WS, Mordi I, Mahrous A, Ahmed N, Wilson R, Lasalle L, Généreux P, Ford I, Berry C (2014) A randomized trial of deferred stenting versus immediate stenting to prevent no- or slow-reflow in acute ST-segment elevation myocardial infarction (DEFER-STEMI). *J Am Coll Cardiol* 63:2088-2098.
- Chappell D, Dörfler N, Jacob M, Rehm M, Welsch U, Conzen P, Becker BF (2010) Glycocalyx protection reduces leukocyte adhesion after ischemia/reperfusion. *Shock* 34:133-139.
- Chappell D, Jacob M, Hofmann-Kiefer K, Bruegger D, Rehm M, Conzen P, Welsch U, Becker BF (2007) Hydrocortisone preserves the vascular barrier by protecting the endothelial glycocalyx. *Anesthesiology* 107:776-784.
- Chappell D, Heindl B, Jacob M, Annecke T, Chen C, Rehm M, Conzen P, Becker BF (2011) Sevoflurane reduces leukocyte and platelet adhesion after ischemia-reperfusion by protecting the endothelial glycocalyx. *Anesthesiology* 115:483-491.
- Chen C, Chappell D, Annecke T, Conzen P, Jacob M, Welsch U, Zwissler B, Becker BF (2016) Sevoflurane mitigates shedding of hyaluronan from the coronary endothelium, also during ischemia/reperfusion: an ex vivo animal study. *Hypoxia (Auckl)* 4:81-90.
- Cheung PY, Sawicki G, Wozniak M, Wang W, Radomski MW, Schulz R (2000) Matrix metalloproteinase-2 contributes to ischemia-reperfusion injury in the heart. *Circulation* 101:1833-1839.
- Chislock EM, Pendergast AM (2013) Abl family kinases regulate endothelial barrier function in vitro and in mice. *PLoS One* 8:e85231.
- Constantinescu AA, Vink H, Spaan JA (2003) Endothelial cell glycocalyx modulates immobilization of leukocytes at the endothelial surface. *Arterioscler Thromb Vasc Biol* 23:1541-1547.
- Culic O, Gruwel ML, Schrader J (1997) Energy turnover of vascular endothelial cells. *The American journal of physiology* 273:C205-213.
- Davis GE, Senger DR (2005) Endothelial extracellular matrix: biosynthesis, remodeling, and functions during vascular morphogenesis and neovessel stabilization. *Circ Res* 97:1093-1107.
- de Waha S, Patel MR, Granger CB, Ohman EM, Maehara A, Eitel I, Ben-Yehuda O, Jenkins P, Thiele H, Stone GW (2017) Relationship between microvascular obstruction and adverse events following primary percutaneous coronary intervention for ST-segment elevation myocardial infarction: an individual patient data pooled analysis from seven randomized trials. *Eur Heart J* 38:3502-3510.
- Dehghani T, Thai PN, Sodhi H, Ren L, Sirish P, Nader CE, Timofeyev V, Overton JL, Li X, Lam KS, Chiamvimonvat N, Panitch A (2022) Selectin-targeting glycosaminoglycan-peptide conjugate limits neutrophil-mediated cardiac reperfusion injury. *Cardiovasc Res* 118:267-281.
- Dejana E (2004) Endothelial cell-cell junctions: happy together. *Nature reviews Molecular cell biology* 5:261-270.
- Dejana E, Orsenigo F, Lampugnani MG (2008) The role of adherens junctions and VE-cadherin in the control of vascular permeability. *J Cell Sci* 121:2115-2122.
- Desmet W, Bogaert J, Dubois C, Sinnaeve P, Adriaenssens T, Pappas C, Ganame J, Dymarkowski S, Janssens S, Belmans A, Van de Werf F (2011) High-dose intracoronary adenosine for myocardial salvage in patients with acute ST-segment elevation myocardial infarction. *Eur Heart J* 32:867-877.
- Esser S, Lampugnani MG, Corada M, Dejana E, Risau W (1998) Vascular endothelial growth factor induces VE-cadherin tyrosine phosphorylation in endothelial cells. *J Cell Sci* 111 ( Pt 13):1853-1865.
- Ferré-Vallverdú M, Sánchez-Lacuesta E, Plaza-López D, Diez-Gil JL, Sepúlveda-Sanchis P, Gil-Cayuela C, Maceira-Gonzalez A, Miró-Palau V, Montero-Argudo A, Martínez-Dolz L, Igual-Muñoz B (2021) Prognostic value and clinical predictors of intramyocardial hemorrhage measured by CMR T2\* sequences in STEMI. *Int J Cardiovasc Imaging* 37:1735-1744.
- Ferrera R, Benhabbouche S, Da Silva CC, Alam MR, Ovize M (2015) Delayed low pressure at reperfusion: A new approach for cardioprotection. *J Thorac Cardiovasc Surg* 150:1641-1648.e1642.
- Fonseca D, Antunes P, Cotrim M (2016) The Morphology, Physiology and Pathophysiology of Coronary Microcirculation. In, pp 15-47.
- Galaup A, Gomez E, Souktani R, Durand M, Cazes A, Monnot C, Teillon J, Le Jan S, Bouleti C, Briois G, Philippe J, Pons S, Martin V, Assaly R, Bonnin P, Ratajczak P, Janin A, Thurston G, Valenzuela DM, Murphy AJ, Yancopoulos GD, Tissier R, Berdeaux A, Ghaleh B, Germain S (2012) Protection against myocardial infarction and no-reflow through preservation of vascular integrity by angiopoietin-

- like 4. *Circulation* 125:140-149.
- Gao X, Kouklis P, Xu N, Minshall RD, Sandoval R, Vogel SM, Malik AB (2000) Reversibility of increased microvessel permeability in response to VE-cadherin disassembly. *Am J Physiol Lung Cell Mol Physiol* 279:L1218-1225.
- Gao XM, Su Y, Moore S, Han LP, Kiriazis H, Lu Q, Zhao WB, Ruze A, Fang BB, Duan MJ, Du XJ (2019) Relaxin mitigates microvascular damage and inflammation following cardiac ischemia-reperfusion. *Basic Res Cardiol* 114:30.
- Garrido-Urbani S, Bradfield PF, Imhof BA (2014) Tight junction dynamics: the role of junctional adhesion molecules (JAMs). *Cell and tissue research* 355:701-715.
- Green DJ, Hopman MT, Padilla J, Laughlin MH, Thijssen DH (2017) Vascular Adaptation to Exercise in Humans: Role of Hemodynamic Stimuli. *Physiological reviews* 97:495-528.
- Hamid T, Xu Y, Ismahil MA, Rokosh G, Jinno M, Zhou G, Wang Q, Prabhu SD (2022) Cardiac Mesenchymal Stem Cells Promote Fibrosis and Remodeling in Heart Failure: Role of PDGF Signaling. *JACC Basic Transl Sci* 7:465-483.
- Heusch G (2017) Critical Issues for the Translation of Cardioprotection. *Circ Res* 120:1477-1486.
- Heusch G, Skyschally A, Schulz R (2011) The in-situ pig heart with regional ischemia/reperfusion - ready for translation. *J Mol Cell Cardiol* 50:951-963.
- Hollander MR, de Waard GA, Konijnenberg LS, Meijer-van Putten RM, van den Brom CE, Paauw N, de Vries HE, van de Ven PM, Aman J, Van Nieuw-Amerongen GP, Hordijk PL, Niessen HW, Horrevoets AJ, Van Royen N (2016) Dissecting the Effects of Ischemia and Reperfusion on the Coronary Microcirculation in a Rat Model of Acute Myocardial Infarction. *PLoS One* 11:e0157233.
- Hu Y-J, Wang Y-D, Tan F-Q, Yang W-XJMbr (2013) Regulation of paracellular permeability: factors and mechanisms. *40:6123-6142*.
- Huang Y, Lei D, Chen Z, Xu B (2021) Factors associated with microvascular occlusion in patients with ST elevation myocardial infarction after primary percutaneous coronary intervention. *J Int Med Res* 49:3000605211024490.
- Husser O, Bodi V, Sanchis J, Nunez J, Lopez-Lereu MP, Monmeneu JV, Gomez C, Rumiz E, Merlos P, Bonanad C, Minana G, Valero E, Chaustre F, Forteza MJ, Rieger GA, Chorro FJ, Llacer A (2013) Predictors of cardiovascular magnetic resonance-derived microvascular obstruction on patient admission in STEMI. *Int J Cardiol* 166:77-84.
- Ibanez B, James S, Agewall S, Antunes MJ, Bucciarelli-Ducci C, Bueno H, Caforio ALP, Crea F, Goudevenos JA, Halvorsen S, Hindricks G, Kastrati A, Lenzen MJ, Prescott E, Roffi M, Valgimigli M, Varenhorst C, Vranckx P, Widimský P (2018) 2017 ESC Guidelines for the management of acute myocardial infarction in patients presenting with ST-segment elevation: The Task Force for the management of acute myocardial infarction in patients presenting with ST-segment elevation of the European Society of Cardiology (ESC). *Eur Heart J* 39:119-177.
- Jayadev R, Sherwood DR (2017) Basement membranes. *Curr Biol* 27:R207-r211.
- Jensen CJ, Eberle HC, Nassenstein K, Schlosser T, Farazandeh M, Naber CK, Sabin GV, Bruder O (2011) Impact of hyperglycemia at admission in patients with acute ST-segment elevation myocardial infarction as assessed by contrast-enhanced MRI. *Clin Res Cardiol* 100:649-659.
- Kalogeris T, Bao Y, Korthuis RJ (2014) Mitochondrial reactive oxygen species: a double edged sword in ischemia/reperfusion vs preconditioning. *Redox biology* 2:702-714.
- Kasseckert SA, Schäfer C, Kluger A, Gligorievski D, Tillmann J, Schlüter KD, Noll T, Sauer H, Piper HM, Abdallah Y (2009) Stimulation of cGMP signalling protects coronary endothelium against reperfusion-induced intercellular gap formation. *Cardiovasc Res* 83:381-387.
- Kelbæk H, Høfsten DE, Køber L, Helqvist S, Kløvgård L, Holmvang L, Jørgensen E, Pedersen F, Saunamäki K, De Backer O, Bang LE, Kofeod KF, Lønborg J, Ahtarovski K, Vejlsstrup N, Bøtker HE, Terkelsen CJ, Christiansen EH, Ravkilde J, Tilsted HH, Villadsen AB, Aarøe J, Jensen SE, Raungaard B, Jensen LO, Clemmensen P, Grande P, Madsen JK, Torp-Pedersen C, Engstrøm T (2016) Deferred versus conventional stent implantation in patients with ST-segment elevation myocardial infarction (DANAMI 3-DEFER): an open-label, randomised controlled trial. *Lancet* 387:2199-2206.
- Kleinbongard P, Bøtker HE, Ovize M, Hausenloy DJ, Heusch G (2020) Co-morbidities and co-medications as confounders of cardioprotection-Does it matter in the clinical setting? *British journal of pharmacology* 177:5252-5269.
- Kloner RA, Ganote CE, Jennings RB (1974) The "no-reflow" phenomenon after temporary coronary occlusion in the dog. *The Journal of clinical investigation* 54:1496-1508.
- Komarova YA, Kruse K, Mehta D, Malik AB (2017) Protein Interactions at Endothelial Junctions and Signaling Mechanisms Regulating Endothelial Permeability. *Circ Res* 120:179-206.
- Konijnenberg LSF, Damman P, Duncker DJ, Kloner RA, Nijveldt R, van Geuns RM, Berry C, Riksen NP, Escaned J, van Royen N (2020) Pathophysiology and diagnosis of coronary microvascular dysfunction in ST-elevation myocardial infarction. *Cardiovasc Res* 116:787-805.
- Koning NJ, de Lange F, van Meurs M, Jongman RM, Ahmed Y, Schwarte LA, van Nieuw Amerongen GP, Vonk ABA, Niessen HW, Baufreton C, Boer C (2018) Reduction of vascular leakage by imatinib is associated with preserved microcirculatory perfusion and reduced renal injury markers in a rat model of cardiopulmonary bypass. *Br J Anaesth* 120:1165-1175.
- Krzywonos-Zawadzka A, Franczak A, Olejnik A, Radomski M, Gilmer JF, Sawicki G, Woźniak M, Bil-Lula I (2019) Cardioprotective effect of MMP-2-inhibitor-NO-donor hybrid against ischaemia/reperfusion injury. *Journal of cellular and molecular medicine* 23:2836-2848.
- Landmesser U, Dikalov S, Price SR, McCann L, Fukui T, Holland SM, Mitch WE, Harrison DG (2003) Oxidation of tetrahydrobiopterin leads to uncoupling of endothelial cell nitric oxide synthase in hypertension. *The Journal of clinical investigation* 111:1201-1209.
- Le TL, Yap AS, Stow JL (1999) Recycling of E-cadherin: a potential mechanism for regulating cadherin dynamics. *The Journal of cell biology* 146:219-232.
- Lecour S, Andreadou I, Bøtker HE, Davidson SM, Heusch G, Ruiz-Meana M, Schulz R, Zuurbier CJ, Ferdinandy P, Hausenloy DJ (2021) Improving Preclinical Assessment of Cardioprotective Therapies (IMPACT) criteria: guidelines of the EU-CARDIOPROTECTION COST Action. *Basic Res Cardiol* 116:52.
- Lee SJ, Lee CK, Kang S, Park I, Kim YH, Kim SK, Hong SP, Bae H, He Y, Kubota Y, Koh GY (2018) Angiotensin-2 exacerbates cardiac hypoxia and inflammation after myocardial infarction. *The Journal of clinical investigation* 128:5018-5033.
- Lee SW, Won JY, Lee HY, Lee HJ, Youn SW, Lee JY, Cho CH, Cho HJ, Oh S, Chae IH, Kim HS (2011) Angiotensin-1 protects heart against ischemia/reperfusion injury through VE-cadherin dephosphorylation and myocardial integrin-β1/ERK/caspase-9 phosphorylation cascade. *Molecular medicine (Cambridge, Mass)* 17:1095-1106.

- Li C, Ma Q, Toan S, Wang J, Zhou H, Liang J (2020) SERCA overexpression reduces reperfusion-mediated cardiac microvascular damage through inhibition of the calcium/MCU/mPTP/necroptosis signaling pathways. *Redox biology* 36:101659.
- Li S, Chen J, Liu M, Chen Y, Wu Y, Li Q, Ma T, Gao J, Xia Y, Fan M, Chen A, Lu D, Su E, Xu F, Chen Z, Qian J, Ge J (2021) Protective effect of HINT2 on mitochondrial function via repressing MCU complex activation attenuates cardiac microvascular ischemia-reperfusion injury. *Basic Res Cardiol* 116:65.
- Li Y, Yao Y, Li J, Chen Q, Zhang L, Wang QK (2019) Losartan protects against myocardial ischemia and reperfusion injury via vascular integrity preservation. *Faseb j* 33:8555-8564.
- Lindsey ML, Bolli R, Canty JM, Jr., Du XJ, Frangogiannis NG, Frantz S, Gourdie RG, Holmes JW, Jones SP, Kloner RA, Lefler DJ, Liao R, Murphy E, Ping P, Przyklenk K, Recchia FA, Schwartz Longacre L, Ripplinger CM, Van Eyk JE, Heusch G (2018) Guidelines for experimental models of myocardial ischemia and infarction. *Am J Physiol Heart Circ Physiol* 314:H812-h838.
- Liu T, Howarth AG, Chen Y, Nair AR, Yang HJ, Ren D, Tang R, Sykes J, Kovacs MS, Dey D, Slomka P, Wood JC, Finney R, Zeng M, Prato FS, Francis J, Berman DS, Shah PK, Kumar A, Dharmakumar R (2022) Intramyocardial Hemorrhage and the "Wave Front" of Reperfusion Injury Compromising Myocardial Salvage. *J Am Coll Cardiol* 79:35-48.
- Machado LS, Kozak A, Ergul A, Hess DC, Borlongan CV, Fagan SC (2006) Delayed minocycline inhibits ischemia-activated matrix metalloproteinases 2 and 9 after experimental stroke. *BMC Neurosci* 7:56.
- Magruder JT, Grimm JC, Crawford TC, Johnston L, Santhanam L, Stephens RS, Berkowitz DE, Shah AS, Bush EL, Damarla M, Damico RL, Hassoun PM, Kim BS (2018) Imatinib Is Protective Against Ischemia-Reperfusion Injury in an Ex Vivo Rabbit Model of Lung Injury. *Ann Thorac Surg* 105:950-956.
- Maier CM, Hsieh L, Crandall T, Narasimhan P, Chan PH (2006) Evaluating therapeutic targets for reperfusion-related brain hemorrhage. *Ann Neurol* 59:929-938.
- Maxwell L, Gavin JB (1991) The role of post-ischaemic reperfusion in the development of microvascular incompetence and ultrastructural damage in the myocardium. *Basic Res Cardiol* 86:544-553.
- Merali Z, Leung J, Mikulis D, Silver F, Kassner A (2015) Longitudinal assessment of imatinib's effect on the blood-brain barrier after ischemia/reperfusion injury with permeability MRI. *Transl Stroke Res* 6:39-49.
- Mertens S, Noll T, Spahr R, Krützfeldt A, Piper HM (1990) Energetic response of coronary endothelial cells to hypoxia. *The American journal of physiology* 258:H689-694.
- Musiolic J, van Caster P, Skyschally A, Boengler K, Gres P, Schulz R, Heusch G (2010) Reduction of infarct size by gentle reperfusion without activation of reperfusion injury salvage kinases in pigs. *Cardiovasc Res* 85:110-117.
- Nair AR, Johnson EA, Yang HJ, Cokic I, Francis J, Dharmakumar R (2020) Reperfused hemorrhagic myocardial infarction in rats. *PLoS One* 15:e0243207.
- Nazir SA, McCann GP, Greenwood JP, Kunadian V, Khan JN, Mahmoud IZ, Blackman DJ, Been M, Abrams KR, Shipley L, Wilcox R, Adgey AA, Gershlick AH (2016) Strategies to attenuate micro-vascular obstruction during P-PCI: the randomized reperfusion facilitated by local adjunctive therapy in ST-elevation myocardial infarction trial. *Eur Heart J* 37:1910-1919.
- Nieuwdorp M, Meuwese MC, Mooij HL, Ince C, Broekhuizen LN, Kastelein JJ, Stroes ES, Vink H (2008) Measuring endothelial glycocalyx dimensions in humans: a potential novel tool to monitor vascular vulnerability. *J Appl Physiol* (1985) 104:845-852.
- Nitta T, Hata M, Gotoh S, Seo Y, Sasaki H, Hashimoto N, Furuse M, Tsukita S (2003) Size-selective loosening of the blood-brain barrier in claudin-5-deficient mice. *The Journal of cell biology* 161:653-660.
- O'Donnell VB, Chumley PH, Hogg N, Bloodsworth A, Darley-Usmar VM, Freeman BA (1997) Nitric oxide inhibition of lipid peroxidation: kinetics of reaction with lipid peroxy radicals and comparison with alpha-tocopherol. *Biochemistry* 36:15216-15223.
- Okamoto F, Allen BS, Buckberg GD, Bugyi H, Leaf J (1986) Reperfusion conditions: importance of ensuring gentle versus sudden reperfusion during relief of coronary occlusion. *J Thorac Cardiovasc Surg* 92:613-620.
- Orsenigo F, Giampietro C, Ferrari A, Corada M, Galaup A, Sigismund S, Ristagno G, Maddaluno L, Koh GY, Franco D, Kurtcuoglu V, Poulidakos D, Baluk P, McDonald D, Grazia Lampugnani M, Dejana E (2012) Phosphorylation of VE-cadherin is modulated by haemodynamic forces and contributes to the regulation of vascular permeability in vivo. *Nat Commun* 3:1208.
- Ota S, Nishiguchi T, Taruya A, Tanimoto T, Ino Y, Katayama Y, Ozaki Y, Satogami K, Tanaka A (2022) Hyperglycemia and intramyocardial hemorrhage in patients with ST-segment elevation myocardial infarction. *J Cardiol* 10.1016/j.jjcc.2022.06.003.
- Pahakis MY, Kosky JR, Dull RO, Tarbell JM (2007) The role of endothelial glycocalyx components in mechanotransduction of fluid shear stress. *Biochem Biophys Res Commun* 355:228-233.
- Passov A, Schramko A, Salminen US, Aittomäki J, Andersson S, Pesonen E (2021) Endothelial glycocalyx during early reperfusion in patients undergoing cardiac surgery. *PLoS One* 16:e0251747.
- Peterson JT, Li H, Dillon L, Bryant JW (2000) Evolution of matrix metalloprotease and tissue inhibitor expression during heart failure progression in the infarcted rat. *Cardiovasc Res* 46:307-315.
- Pike MM, Luo CS, Clark MD, Kirk KA, Kitakaze M, Madden MC, Cragoe EJ, Jr., Pohost GM (1993) NMR measurements of Na<sup>+</sup> and cellular energy in ischemic rat heart: role of Na<sup>(+)</sup>-H<sup>+</sup> exchange. *The American journal of physiology* 265:H2017-2026.
- Platts SH, Linden J, Duling BR (2003) Rapid modification of the glycocalyx caused by ischemia-reperfusion is inhibited by adenosine A2A receptor activation. *Am J Physiol Heart Circ Physiol* 284:H2360-2367.
- Potter MD, Barbero S, Cheresch DA (2005) Tyrosine phosphorylation of VE-cadherin prevents binding of p120- and beta-catenin and maintains the cellular mesenchymal state. *The Journal of biological chemistry* 280:31906-31912.
- Querio G, Antoniotti S, Geddo F, Tullio F, Penna C, Pagliaro P, Gallo MP (2021) Ischemic heart disease and cardioprotection: Focus on estrogenic hormonal setting and microvascular health. *Vascular pharmacology* 141:106921.
- Quintero M, Colombo SL, Godfrey A, Moncada S (2006) Mitochondria as signaling organelles in the vascular endothelium. *Proceedings of the National Academy of Sciences of the United States of America* 103:5379-5384.
- Ramnath R, Foster RR, Qiu Y, Cope G, Butler MJ, Salmon AH, Mathieson PW, Coward RJ, Welsh GI, Satchell SC (2014) Matrix metalloproteinase 9-mediated shedding

- of syndecan 4 in response to tumor necrosis factor  $\alpha$ : a contributor to endothelial cell glycocalyx dysfunction. *Faseb j* 28:4686-4699.
- Reffellmann T, Hale SL, Li G, Kloner RA (2002) Relationship between no reflow and infarct size as influenced by the duration of ischemia and reperfusion. *J American Journal of Physiology-Heart Circ Physiol* 282:H766-H772.
- Rehm M, Zahler S, Lötsch M, Welsch U, Conzen P, Jacob M, Becker BF (2004) Endothelial glycocalyx as an additional barrier determining extravasation of 6% hydroxyethyl starch or 5% albumin solutions in the coronary vascular bed. *Anesthesiology* 100:1211-1223.
- Reimer KA, Lowe JE, Rasmussen MM, Jennings RB (1977) The wavefront phenomenon of ischemic cell death. 1. Myocardial infarct size vs duration of coronary occlusion in dogs. *Circulation* 56:786-794.
- Reinstadler SJ, Stiermaier T, Reindl M, Feistritzer HJ, Fuernau G, Eitel C, Desch S, Klug G, Thiele H, Metzler B, Eitel I (2019) Intramyocardial haemorrhage and prognosis after ST-elevation myocardial infarction. *Eur Heart J Cardiovasc Imaging* 20:138-146.
- Reitsma S, Slaaf DW, Vink H, van Zandvoort MA, oude Egbrink MG (2007) The endothelial glycocalyx: composition, functions, and visualization. *Pflugers Archiv : European journal of physiology* 454:345-359.
- Robbers LF, Eerenberg ES, Teunissen PF, Jansen MF, Hollander MR, Horrevoets AJ, Knaapen P, Nijveldt R, Heymans MW, Levi MM, van Rossum AC, Niessen HW, Marcu CB, Beek AM, van Royen N (2013) Magnetic resonance imaging-defined areas of microvascular obstruction after acute myocardial infarction represent microvascular destruction and haemorrhage. *Eur Heart J* 34:2346-2353.
- Rodrigo R, Libuy M, Feliú F, Hasson D (2013) Molecular basis of cardioprotective effect of antioxidant vitamins in myocardial infarction. *BioMed research international* 27.
- Rodrigo R, Retamal C, Schupper D, Vergara-Hernández D, Saha S, Profumo E, Buttari B, Saso L (2022) Antioxidant Cardioprotection against Reperfusion Injury: Potential Therapeutic Roles of Resveratrol and Quercetin. *Molecules* 27.
- Romancic AM, Harrison SM, Bao W, Burns-Kurtis CL, Pickering S, Gu J, Grau E, Mao J, Sathe GM, Ohlstein EH, Yue TL (2002) Myocardial protection from ischemia/reperfusion injury by targeted deletion of matrix metalloproteinase-9. *Cardiovasc Res* 54:549-558.
- Roos ST, Timmers L, Biesbroek PS, Nijveldt R, Kamp O, van Rossum AC, van Hout GP, Stella PR, Doevendans PA, Knaapen P, Velthuis BK, van Royen N, Voskuil M, Nap A, Appelman Y (2016) No benefit of additional treatment with exenatide in patients with an acute myocardial infarction. *Int J Cardiol* 220:809-814.
- Rubio-Gayosso I, Platts SH, Duling BR (2006) Reactive oxygen species mediate modification of glycocalyx during ischemia-reperfusion injury. *Am J Physiol Heart Circ Physiol* 290:H2247-2256.
- Saleem S, Wang D, Zhao T, Sullivan RD, Reed GL (2021) Matrix Metalloproteinase-9 Expression is Enhanced by Ischemia and Tissue Plasminogen Activator and Induces Hemorrhage, Disability and Mortality in Experimental Stroke. *Neuroscience* 460:120-129.
- Sato H, Jordan JE, Zhao ZQ, Sarvotham SS, Vinten-Johansen J (1997) Gradual reperfusion reduces infarct size and endothelial injury but augments neutrophil accumulation. *Ann Thorac Surg* 64:1099-1107.
- Schmidt EP, Yang Y, Janssen WJ, Gandjeva A, Perez MJ, Barthel L, Zemans RL, Bowman JC, Koyanagi DE, Yunt ZX, Smith LP, Cheng SS, Overdier KH, Thompson KR, Geraci MW, Douglas IS, Pearse DB, Tudor RM (2012) The pulmonary endothelial glycocalyx regulates neutrophil adhesion and lung injury during experimental sepsis. *Nat Med* 18:1217-1223.
- Sezer M, Escaned J, Broyd CJ, Umman B, Bugra Z, Ozcan I, Sonsoz MR, Ozcan A, Atici A, Aslanger E, Sezer ZI, Davies JE, van Royen N, Umman S (2022) Gradual Versus Abrupt Reperfusion During Primary Percutaneous Coronary Interventions in ST-Segment-Elevation Myocardial Infarction (GUARD). *J Am Heart Assoc* 11:e024172.
- Su EJ, Fredriksson L, Geyer M, Folestad E, Cale J, Andrae J, Gao Y, Pietras K, Mann K, Yepes M, Strickland DK, Betsholtz C, Eriksson U, Lawrence DA (2008) Activation of PDGF-CC by tissue plasminogen activator impairs blood-brain barrier integrity during ischemic stroke. *Nat Med* 14:731-737.
- Sumii T, Lo EH (2002) Involvement of matrix metalloproteinase in thrombolysis-associated hemorrhagic transformation after embolic focal ischemia in rats. *Stroke* 33:831-836.
- Takeo S, Liu JX, Tanonaka K, Nasa Y, Yabe K, Tanahashi H, Sudo H (1995) Reperfusion at reduced flow rates enhances postischemic contractile recovery of perfused heart. *The American journal of physiology* 268:H2384-2395.
- Tanaka S, Chen-Yoshikawa TF, Kajiwaru M, Menju T, Ohata K, Takahashi M, Kondo T, Hijiyama K, Motoyama H, Aoyama A, Masuda S, Date H (2016) Protective Effects of Imatinib on Ischemia/Reperfusion Injury in Rat Lung. *Ann Thorac Surg* 102:1717-1724.
- van den Berg BM, Vink H, Spaan JA (2003) The endothelial glycocalyx protects against myocardial edema. *Circ Res* 92:592-594.
- van Haaren PM, VanBavel E, Vink H, Spaan JA (2003) Localization of the permeability barrier to solutes in isolated arteries by confocal microscopy. *Am J Physiol Heart Circ Physiol* 285:H2848-2856.
- van Kranenburg M, Magro M, Thiele H, de Waha S, Eitel I, Cochet A, Cottin Y, Atar D, Buser P, Wu E, Lee D, Bodi V, Klug G, Metzler B, Delewi R, Bernhardt P, Rottbauer W, Boersma E, Zijlstra F, van Geuns RJ (2014) Prognostic value of microvascular obstruction and infarct size, as measured by CMR in STEMI patients. *JACC Cardiovasc Imaging* 7:930-939.
- Vijayalakshmi K, Whittaker VJ, Kunadian B, Graham J, Wright RA, Hall JA, Sutton A, de Belder MA (2006) Prospective, randomised, controlled trial to study the effect of intracoronary injection of verapamil and adenosine on coronary blood flow during percutaneous coronary intervention in patients with acute coronary syndromes. *Heart* 92:1278-1284.
- Vink H, Duling BR (2000) Capillary endothelial surface layer selectively reduces plasma solute distribution volume. *Am J Physiol Heart Circ Physiol* 278:H285-289.
- Wang G, Kostidis S, Tiemeier GL, Sol W, de Vries MR, Giera M, Carmeliet P, van den Berg BM, Rabelink TJ (2020) Shear Stress Regulation of Endothelial Glycocalyx Structure Is Determined by Glucobiosynthesis. *Arterioscler Thromb Vasc Biol* 40:350-364.
- Wessel F, Winderlich M, Holm M, Frye M, Rivera-Galdos R, Vockel M, Linnepe R, Ipe U, Stadtmann A, Zarbock A, Nottebaum AF, Vestweber D (2014) Leukocyte extravasation and vascular permeability are each controlled in vivo by different tyrosine residues of VE-cadherin. *Nature immunology* 15:223-230.
- Whitman GJ, Niibori K, Yokoyama H, Crestanello JA, Lingle DM, Momeni R (1997) The mechanisms of coenzyme Q10 as therapy for myocardial ischemia reperfusion injury. *Mol Aspects Med* 18 Suppl:S195-203.
- Wiesinger A, Peters W, Chappell D, Kentrup D, Reuter

- S, Pavenstädt H, Oberleithner H, Kümpers P (2013) Nanomechanics of the endothelial glycocalyx in experimental sepsis. *PLoS One* 8:e80905.
- Yu H, Kalogeris T, Korthuis RJ (2019) Reactive species-induced microvascular dysfunction in ischemia/reperfusion. *Free radical biology & medicine* 135:182-197.
- Zhou T, Chuang CC, Zuo L (2015) Molecular Characterization of Reactive Oxygen Species in Myocardial Ischemia-Reperfusion Injury. *BioMed research international* 2015:864946.